

Three New Compounds from the Aerial Parts of *Caragana sinica*

by Shuguang Wang, Dayou Ma, and Changqi Hu*

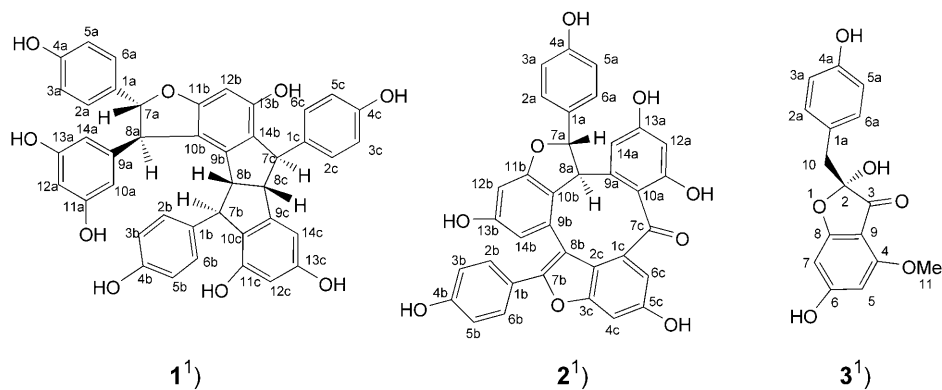
Department of Chemistry of Natural Drugs, School of Pharmacy, Fudan University, Shanghai 200032, People's Republic of China
(phone: + 86-021-54237564; e-mail: shuguang3@shmu.edu.cn)

Two new stilbenoids, carasiphenol C (**1**) and Carasiphenol D (**2**), and one new dihydroaurone, carasinaurone (**3**), were isolated from the aerial parts of *Caragana sinica*, along with three known substances. Their structures were elucidated on the basis of spectroscopic methods and X-ray-diffraction analysis. Compound **2** showed significant biological activity against *Pyricularia oryzae*.

Introduction. – *Caragana sinica* (BUCHOLZ.) REHD. (Leguminosae) is widely distributed in China. Its dried root named 'Jinquen' [1] has been used as a folk medicine. In the previous papers, we reported that the AcOEt extract of the root of *Caragana sinica* showed the effects of stimulating the proliferation, differentiation, and maturation of cultured osteoblasts *in vitro* [2]. In our lab, many oligostilbenes, some of which exhibited estrogenic activity, were isolated from this plant [3]. The aerial parts of the plant have scarcely been used and were always considered a waste product. However, our recent studies on the aerial parts of the plant showed that it can be a rich source of oligostilbenes. In this paper, we report the isolation and structure elucidation of two new oligostilbenes, carasiphenol C (**1**) and carasiphenol D (**2**), and a new dihydroaurone, carasinaurone (**3**), together with three known compounds, (–)-ampelopsin F, caraphenol A [4], and carasinols A [5], from the aerial parts of *Caragana sinica*. Carasiphenol D (**2**) showed significant activity against the mycelial growth of *Pyricularia oryzae*. The minimum inhibitory concentration (MIC) of **2** was 0.017 μM .

Results and Discussion. – Carasiphenol C (**1**) was isolated as a yellowish, amorphous, optically active powder. Its molecular formula, $\text{C}_{42}\text{H}_{32}\text{O}_9$, was deduced from the HR-ESI-MS (m/z 681.2130 ($[M - \text{H}]^+$, $\text{C}_{42}\text{H}_{33}\text{O}_9^+$)). The structure of **2** was established by the ^1H - and ^{13}C -NMR spectra (Tables 1 and 2) and their comparison with those of ampelopsin H [6], a tetramer of resveratrol, and confirmed by HMBC and NOE data (Fig. 1).

The ^1H -NMR spectrum of **1** exhibited signals for three 4-hydroxy-substituted phenyl moieties at δ 7.22 (d , $J = 8.5$ Hz, 2 H), 6.86 (d , $J = 8.5$ Hz, 2 H), 6.98 (d , $J = 8.3$ Hz, 2 H), 6.70 (d , $J = 8.3$ Hz, 2 H), 6.44 (d , $J = 8.6$ Hz, 2 H) and 6.35 (d , $J = 8.3$ Hz, 2 H); for two 3,5-dihydroxy-substituted phenyl moieties at δ 6.25 (d , $J = 2.3$ Hz, 2 H), 6.31 (t , $J = 2.1$ Hz, 1 H), 6.61 (d , $J = 1.3$ Hz, 1 H), and 6.17 (d , $J = 1.3$ Hz, 1 H), and for two aliphatic protons of a dihydrobenzofuran moiety at δ 5.25 ($J = 7.5$ Hz, 1 H) and 4.82 ($J = 7.5$ Hz, 1 H). The ^{13}C -NMR spectrum exhibited the signals of six aliphatic and 36 aromatic C-atoms. All these data suggested that **1** could be a resveratrol trimer.¹ ^1H - and ^{13}C -NMR features of **1** were similar to those of a segment of ampelopsin H [6], except that ampelopsin H was a tetramer. The planar structure of **1** was determined by a HMBC

Table 1. ¹H-NMR Data ((D₆)Acetone, 500 MHz) for **1–3¹⁾**. δ in ppm, J in Hz.

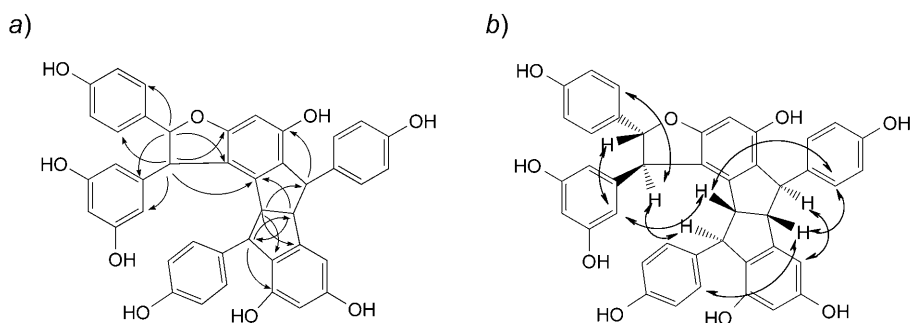
	1	2	3	
H–C(2), H–C(6a)	7.22 (<i>d</i> , <i>J</i> = 8.5)	7.11 (<i>d</i> , <i>J</i> = 8.2)	H–C(5)	5.91 (<i>d</i> , <i>J</i> = 1.3)
H–C(3a), H–C(5a)	6.86 (<i>d</i> , <i>J</i> = 8.5)	6.72 (<i>d</i> , <i>J</i> = 8.6)	H–C(7)	5.97 (<i>d</i> , <i>J</i> = 1.3)
H–C(7a)	5.25 (<i>d</i> , <i>J</i> = 7.5)	5.98 (<i>s</i>)	H–C(10)	3.01 (<i>s</i>)
H–C(8a)	4.82 (<i>d</i> , <i>J</i> = 7.5)	4.81 (<i>s</i>)	H–C(11)	3.74 (<i>s</i>)
H–C(10a)	6.25 (<i>d</i> , <i>J</i> = 2.3)		H–C(2b)	7.02 (<i>d</i> , <i>J</i> = 8.3)
H–C(12a)	6.31 (<i>d</i> , <i>J</i> = 2.1)	6.79 (<i>d</i> , <i>J</i> = 1.9)	H–C(6b)	7.02 (<i>d</i> , <i>J</i> = 8.3)
H–C(14a)	6.25 (<i>d</i> , <i>J</i> = 2.3)	6.85 (<i>d</i> , <i>J</i> = 1.9)	H–C(3b)	6.62 (<i>d</i> , <i>J</i> = 8.5)
H–C(2b), H–C(6b)	6.44 (<i>d</i> , <i>J</i> = 8.6)	7.57 (<i>d</i> , <i>J</i> = 8.7)	H–C(5b)	6.62 (<i>d</i> , <i>J</i> = 8.5)
H–C(3b), H–C(5b)	6.35 (<i>d</i> , <i>J</i> = 8.4)	6.83 (<i>d</i> , <i>J</i> = 8.7)		
H–C(7b)	4.51 (<i>br. s</i>)			
H–C(8b)	3.66 (<i>d</i> , <i>J</i> = 5.9)			
H–C(12b)	6.25 (<i>s</i>)	6.50 (<i>d</i> , <i>J</i> = 2.1)		
H–C(14b)		6.25 (<i>d</i> , <i>J</i> = 2.1)		
H–C(2c)	6.98 (<i>d</i> , <i>J</i> = 8.3)			
H–C(6c)	6.98 (<i>d</i> , <i>J</i> = 8.3)	6.66 (<i>d</i> , <i>J</i> = 2.1)		
H–C(4c)		6.44 (<i>d</i> , <i>J</i> = 2.1)		
H–C(3c)	6.70 (<i>d</i> , <i>J</i> = 8.3)			
H–C(5c)	6.70 (<i>d</i> , <i>J</i> = 8.3)			
H–C(7c)	4.61 (<i>br. s</i>)			
H–C(8c)	3.61 (<i>d</i> , <i>J</i> = 5.9)			
H–C(12c)	6.17 (<i>d</i> , <i>J</i> = 1.3)			
H–C(14c)	6.61 (<i>d</i> , <i>J</i> = 1.3)			

experiment (Fig. 1, a). Significant correlations H–C(8b)/C(7c) and C(9c), H–C(8c)/C(9b) and C(7b), H–C(7c)/C(9b) and C(8b) indicated the presence of a five-membered ring (denoted by C₁). Significant correlations H–C(7b)/C(9c), H–C(7c)/C(8c), H–C(8b)/C(9c), H–C(8b)/C(10c), and H–C(8c)/C(7b) indicated the presence of another five-membered ring (noted by C₂), and the two rings C₁ and C₂ were connected at C(8b) and C(8c). This kind of connection is the same as that observed in pallidol [7]. Significant correlations H–C(8a)/C(9b) and C(11b) indicated that C(8a) was connected to C(10b). All above data confirmed the planar structure of **1** as shown in Fig. 1, a. The NOEs H–C(8a)/H–C(2a,6a) and H–C(7a)/H–C(10a,14a) indicated a *trans* orientation of H–C(7a) and H–C(8a) (Fig. 1, b). Similarly, the NOEs

¹⁾ Arbitrary numbering; for systematic names, see *Exper. Part*.

Table 2. ^{13}C -NMR Data ((D₆)Acetone, 125 MHz) for **1–3**¹). δ in ppm.

	1	2	1	2	3		
C(1a)	133.5	133.4	C(10b)	116.6	124.3		
C(2a), C(6a)	129.6	129.0	C(11b)	163.1	156.9	C(2)	108.1
C(3a), C(5a)	116.5	116.4	C(12b)	97.1	103.5	C(3)	195.3
C(4a)	158.7	158.9	C(13b)	155.6	160.5	C(4)	162.6
C(7a)	95.1	89.3	C(14b)	126.1	109.5	C(5)	95.1
C(8a)	57.4	48.2	C(1c)	138.0	139.1	C(6)	168.7
C(9a)	145.8	139.6	C(2c)	129.4	125.3	C(7)	92.3
C(10a)	107.8	120.3	C(6c)	129.4	107.6	C(8)	173.4
C(11a)	160.3	156.1	C(3c)	116.2	159.9	C(9)	105.6
C(12a)	102.6	97.3	C(5c)	116.2	159.7	C(10)	47.2
C(13a)	160.3	157.3	C(4c)	156.2	102.3	C(11)	58.8
C(14a)	107.8	109.5	C(7c)	53.7	199.7	C(1a)	126.5
C(1b)	137.0	123.4	C(8c)	60.4		C(2a)	132.7
C(2b), (6b)	129.4	129.3	C(9b)	150.7		C(6a)	132.7
C(3b), (5b)	115.6	116.3	C(10c)	123.3		C(3a)	116.4
C(4b)	156.3	158.9	C(11c)	159.6		C(5a)	116.4
C(7b)	50.4	150.8	C(12c)	102.8		C(4a)	157.8
C(8b)	60.3	114.9	C(13c)	155.6			
C(9b)	145.4	135.3	C(14c)	103.6			

Fig. 1. Significant a) HMBC and b) NOE correlations for carasiphenol C (**1**)

$\text{H}-\text{C}(8\text{c})/\text{H}-\text{C}(2\text{c},6\text{c})$ and $\text{H}-\text{C}(7\text{c})/\text{H}-\text{C}(14\text{c})$ were in accord with a *trans* orientation of $\text{H}-\text{C}(7\text{c})$ and $\text{H}-\text{C}(8\text{c})$. Also the NOE interactions $\text{H}-\text{C}(8\text{a})/\text{H}-\text{C}(7\text{b})$, $\text{H}-\text{C}(8\text{b})/\text{H}-\text{C}(10\text{a},14\text{a})$ and $\text{H}-\text{C}(8\text{b})/\text{H}-\text{C}(2\text{b},6\text{b})$ suggested that $\text{H}-\text{C}(8\text{c})$ and $\text{H}-\text{C}(7\text{b})$, as well as $\text{H}-\text{C}(7\text{b})$ and $\text{H}-\text{C}(8\text{b})$ would have a *trans* orientation. The NOE interactions $\text{H}-\text{C}(8\text{b})/\text{H}-\text{C}(2\text{c},6\text{c})$, $\text{H}-\text{C}(8\text{c})/\text{H}-\text{C}(2\text{c},6\text{c})$ and $\text{H}-\text{C}(8\text{c})/\text{H}-\text{C}(2\text{b},6\text{b})$ determined the configuration of $\text{H}-\text{C}(8\text{c})$ and $\text{H}-\text{C}(8\text{b})$ to be *cis*; $\text{H}-\text{C}(7\text{b})$ and $\text{H}-\text{C}(8\text{a})$ were determined to be *cis*.

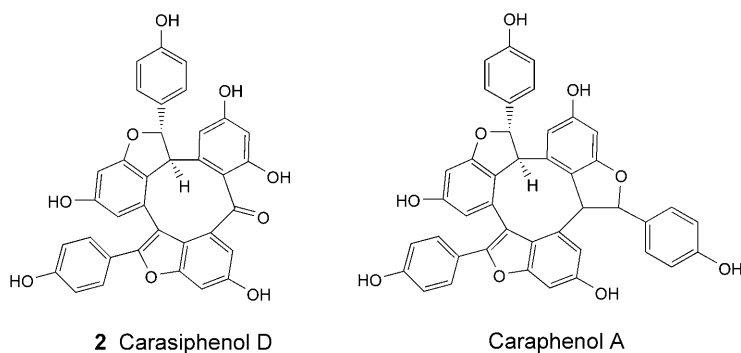
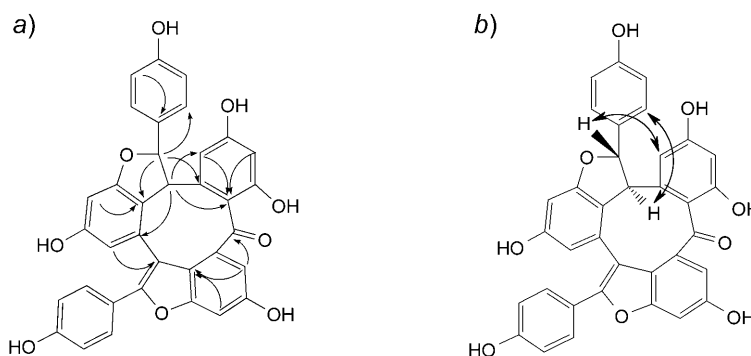
Carasiphenol D (**2**) was isolated as a yellowish amorphous powder. Its molecular formula, $\text{C}_{35}\text{H}_{22}\text{O}_9$, was established by HR-ESI-MS (m/z 587.1318 ($[\text{M} + \text{H}]^+$, $\text{C}_{35}\text{H}_{23}\text{O}_9^+$)). The UV spectrum (λ_{max} 326 nm) revealed the presence of a strongly conjugated system in the structure. The structure of **2** was deduced from its spectral data (Tables 1 and 2), their comparison with those of caraphenol A [4] (Table 3 and Fig. 2), a resveratrol trimer which has also been isolated from the roots of this plant, and confirmed by HMBC and NOESY data (Fig. 3).

Table 3. NMR Data ((D₆)Acetone) of Caraphenol A and Carasiphenol D. δ in ppm.

	Caraphenol A ¹)		Carasiphenol D ¹) (2)	
	δ (H)	δ (C)	δ (H)	δ (C)
C(1a)		133.5		133.4
H–C(2a), H–C(6a)	7.22 (<i>d</i> , <i>J</i> = 8.5)	129.0	7.11 (<i>d</i> , <i>J</i> = 8.2)	129.0
H–C(3a), H–C(5a)	6.86 (<i>d</i> , <i>J</i> = 8.5)	116.5	6.72 (<i>d</i> , <i>J</i> = 8.6)	116.4
C(4a)		158.7		158.9
H–C(7a)	5.25 (<i>d</i> , <i>J</i> = 7.5)	95.1	5.98 (<i>s</i>)	89.3
H–C(8a)	4.82 (<i>d</i> , <i>J</i> = 7.5)	57.4	4.81 (<i>s</i>)	48.2
C(9a)		145.8		139.6
C(10a)		107.8		120.3
C(11a)		160.3		156.1
H–C(12a)	6.31 (<i>d</i> , <i>J</i> = 2.1)	102.6	6.79 (<i>d</i> , <i>J</i> = 1.9)	97.0
C(13a)		160.3		157.3
H–C(14a)	6.25 (<i>d</i> , <i>J</i> = 2.3)	107.8	6.85 (<i>d</i> , <i>J</i> = 1.9)	109.5
C(1b)		122.7		123.4
H–C(2b), H–C(6b)	7.26 (<i>d</i> , <i>J</i> = 8.7)	129.4	7.57 (<i>d</i> , <i>J</i> = 8.7)	129.3
H–C(3b), H–C(5b)	6.70 (<i>d</i> , <i>J</i> = 8.7)	116.2	6.83 (<i>d</i> , <i>J</i> = 8.7)	116.3
C(4b)		156.3		158.9
C(7b)		149.4		150.8
C(8b)		114.5		114.9
C(9b)		135.3		135.3
C(10b)		118.9		124.3
C(11b)		163.5		156.9
H–C(12b)	6.52 (<i>d</i> , <i>J</i> = 2.1)	98.3	6.50 (<i>d</i> , <i>J</i> = 2.1)	103.5
C(13b)		160.7		160.5
H–C(14b)	6.32 (<i>d</i> , <i>J</i> = 2.1)	109.6	6.25 (<i>d</i> , <i>J</i> = 2.1)	109.5
C(1c)		139.7		139.1
C(2c)		120.5		125.3
H–C(6c)	6.94 (<i>d</i> , <i>J</i> = 8.3)	126.1	6.66 (<i>d</i> , <i>J</i> = 2.1)	107.6
C(3c)		155.2		159.9
C(5c)		157.2		159.7
H–C(4c)	6.81 (<i>d</i> , <i>J</i> = 1.8)	96.4	6.44 (<i>d</i> , <i>J</i> = 2.1)	102.3
H–C(7c) or C(7c)	4.87 (<i>br. s</i>)	45.7		199.7
H–C(8c)	5.92 (<i>br. s</i>)	87.9		
C(9c)		132.5		
H–C(10c)	7.05 (<i>d</i> , <i>J</i> = 8.6)	128.3		
H–C(11c)	6.71 (<i>d</i> , <i>J</i> = 8.6)	115.9		
C(12c)		158.2		
H–C(13c)	6.71 (<i>d</i> , <i>J</i> = 8.6)	115.9		
H–C(14c)	7.05 (<i>d</i> , <i>J</i> = 8.6)	128.3		

Carasiphenol D (**2**) was soluble in acetone, AcOEt, MeOH, and dimethylsulfoxide but was insoluble in distilled H₂O. Carasiphenol (**2**) inhibited the mycelial growth of *P. oryzae* by curling and swelling effect (*MIC* = 0.017 μ M), indicating that **2** may act *via* a mechanism similar to the antifungal mechanisms of rhizoxin, which is a commercial fungicidal agent [8]. Therefore, **2** isolated from *C. sinica* may prove to be a valuable antifungal agent against rice-blast disease.

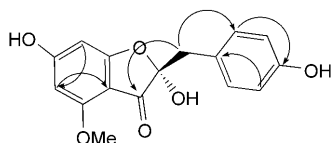
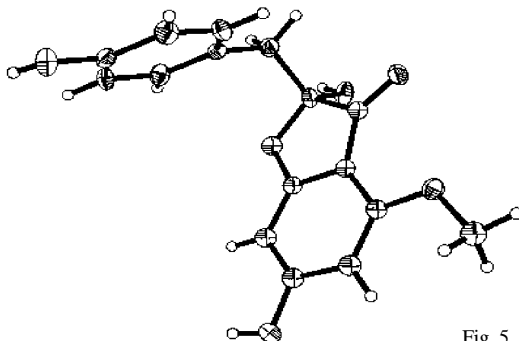
The ¹H-NMR spectrum of **2** exhibited signals for two 4-hydroxy-substituted phenyl groups at δ 7.57 (*d*, *J* = 8.7 Hz, 2 H), 6.83 (*d*, *J* = 8.7 Hz, 2 H), and 7.11 (*d*, *J* = 8.2 Hz, 2 H), and 6.72 (*d*, *J* = 8.6 Hz, 2 H), for three 3,5-dioxy-1,2-phenylene moieties at δ 6.50 (*d*, *J* = 2.1 Hz, 1 H), 6.25 (*d*, *J* = 2.1 Hz, 1 H), 6.44 (*d*, *J* = 2.1 Hz, 1 H),

Fig. 2. Comparison of carasiphenol D (**2**) and caraphenol AFig. 3. Significant a) HMBC and b) NOE correlations for carasiphenol D (**2**)

6.66 (*d*, $J = 2.1$ Hz, 1 H), 6.79 (*d*, $J = 1.9$ Hz, 1 H), and 6.85 (*d*, $J = 1.9$ Hz, 1 H), and for two aliphatic protons of a dihydrobenzofuran moiety at δ 5.98 (br. s, 1 H) and 4.81 (br. s, 1 H). The ^{13}C -NMR spectrum exhibited the signals of two aliphatic and 32 aromatic C-atoms and an isolated carbonyl group (δ 199.7). All these data suggested that **2** could be a stilbene compound. ^1H - and ^{13}C -NMR features of **2** were similar to those of a segment of caraphenol A [4] (Table 3), a resveratrol trimer (Fig. 2).

In the HMBC spectrum of **2**, significant correlations H–C(8a)/C(10a) and C(9b), H–C(14b)/C(8b), H–C(6c)/C(2c) and C(7c), and H–C(14a)/C(10a) indicated the presence of a nine-membered ring (Fig. 3, a). The relative configuration of **2** was established on the basis of the ^1H -NMR and NOESY data. In the ^1H -NMR spectrum, the coupling constant $J(7a,8a)$ of *ca.* 0 Hz suggested that the bond angle between H–C(7a) and H–C(8a) is nearly 90° . According to a molecular model of **2**, *trans* orientation of H–C(7a) and H–C(8a) was deduced. Together with the NOEs interactions H–C(8a)/H–C(2a,6a) and H–C(7a)/H–C(14a) (Fig. 3, b), which also indicated a *trans* orientation for H–C(7a) and H–C(8a) [4], the relative configuration of **2** was established.

Carasinaurone (**3**) was isolated as pale yellow cubic crystals. Its molecular formula $\text{C}_{16}\text{H}_{14}\text{O}_6$ was established by HR-ESI-MS (m/z 341.0428 ($[M + K]^+$, $\text{C}_{16}\text{H}_{14}\text{KO}_6^+$)). The structure of **3** was established by spectroscopic means (Tables 1 and 2, and Fig. 4) and comparison with the data of dihydroaurone [9], which has a planar structure, and its configuration was deduced from the X-ray crystallographic analysis (Fig. 5).

Fig. 4. Significant HMBC correlations of carasinaurone (**3**)Fig. 5. X-Ray structure of carasinaurone (**3**)

The $^1\text{H-NMR}$ spectrum exhibited signals for a 4-hydroxy-substituted phenyl group at δ 7.02 (d , $J = 8.3$ Hz, 2 H) and 6.62 (d , $J = 8.5$ Hz, 2 H), for a 5-hydroxy-3-methoxy-1,2-phenylene group at δ 5.91 (d , $J = 1.3$ Hz, 1 H) and 5.97 ($J = 1.3$ Hz, 1 H), for two CH_2 protons at 3.01 (s), and for three Me protons at δ 3.74 (s). In the HMBC spectrum (Fig. 4), significant correlations $\text{H-C}(10)/\text{C}(2\text{b},6\text{b})$, $\text{C}(3)$, and $\text{C}(2)$, $\text{H-C}(5)/\text{C}(7)$ and $\text{C}(9)$, and $\text{H-C}(7)/\text{C}(5)$ and $\text{C}(9)$ were observed, all above data indicating that **3** could be a dihydroaurone derivative [9] with a planar structure.

X-Ray crystallographic analysis of carasinaurone (**3**)¹): The compound crystallized in the monoclinic space group P_{21}/C with molecules of composition $\text{C}_{16}\text{H}_{14}\text{O}_6$ ($Z = 4$); accurate cell constants are: $a = 10.2922$ (14), $b = 9.2173$ (14), $c = 14.173$ (2) Å; $\beta = 90.497$ (3)°; $V = 1344.5$ (3) Å³. All reflections were collected on the CCD area detector diffractometer, MoK_α radiation (λ 0.71073 Å); maximum 2θ value 25.10°; independent reflections 2387, observed reflections 1797 ($|F|^2 \geq 2 \delta |F|^2$) $F(000) = 632$. The structure was resolved by direct methods (SHELXS-97), expanded by using SHELXS-97/2, and refined by full-matrix least-squares calculations. H-Atoms were fixed at calculated positions. The final indices were $R_1 = 0.0436$, $R_w = 0.1099$.

This work was supported by the National Natural Science Foundation of China (Grant No. 30270155).

Experimental Part

General. M.p.: Kofler micro melting-point apparatus; uncorrected. Optical rotations: Jasco P-1020 polarimeter; MeOH solns. UV Spectra: Shimadzu UV-240 spectrophotometer λ_{max} (log ϵ) in nm. IR Spectra: Perkin-Elmer 783 IR spectrophotometer, KBr pellets; in cm^{-1} . NMR Spectra: Bruker DRX-400 spectrometer; δ in ppm rel. to SiMe_4 as internal standard, J in Hz. HR-ESI-MS: AB-QSTAR-Pulsar mass spectrometer; in m/z (rel. %).

Plant Material. The aerial parts of *C. sinica* (BUCHOLZ.) REHD. were collected in September, 2001, from Zhongxiang County, Hubei Province, China, and identified by Professor Zhi-Jian Feng, Department of Biology, Shanghai East China Normal University. A Voucher specimen (CS-HZ-1106) is deposited in the Herbarium of Materia Medica, Department of Chemistry of Natural Drugs, School of Pharmacy, Fudan University, People's Republic of China.

²) CCDC 245803 contains the supplementary crystallographic data for this paper. These data can be obtained free of charge via www.ccdc.cam.ac.uk/data_request/cif from the Cambridge Crystallographic Data Centre.

Extraction and Isolation. The air-dried and powdered aerial parts of *Caragana sinica* (BUCHOLZ.) REHD. (70 kg) were macerated with 95% EtOH at r.t. The solvent was evaporated to yield a residue (5 kg), which was successively partitioned between H₂O and petroleum ether and AcOEt. The AcOEt extract (230 g) was subjected to CC Silica gel (1.5 kg, 200–300 mesh), CHCl₃/MeOH (10:1, 9:1, 4:1, 7:3 and 3:2), MeOH): *Fractions A–F*. *Fr. C* was subjected to CC (silica gel (300–400 mesh), petroleum ether/Et₂O 3:2): caraphenol A (47 mg), (–)-ampelosin F (6.0 g), and carasinaurone (**3**; 93 mg). The latter was purified by crystallization from MeOH/H₂O. From *Fr. D*, carasiphenol D (**2**; 18 mg) and carasinol A (37 mg) were obtained by CC (silica gel, petroleum ether/Et₂O 2:3; and then *Sephadex LH-20* CHCl₃/MeOH 1:3). Carasiphenol C (**1**; 10 mg) was obtained from *Fr. E* by CC (silica gel, petroleum ether/Et₂O 2:3; then reversed-phase silica gel (*RP-18*), MeOH/H₂O 2:3).

Carasiphenol C (= *1-(3,5-Dihydroxyphenyl)-1,2,6,6a,11,11a-hexahydro-2,6,11-tri(4-hydroxyphenyl)indenol[1',2':2,3]indenol[5,8-b]furan-5,8,10-triol*; **1**): Brown amorphous powder. M.p. 216–218°, [α]_D²⁰ = +36.8 (*c* = 0.281, MeOH). UV (MeOH): 278 (3.2). IR (KBr): 3356, 1605, 1532, 1460, 1320, 1205, 1098, 1016, 883. ¹H- and ¹³C-NMR: *Tables 1* and *2*. HR-ESI-MS: 681.2130 ([*M* + H]⁺, C₄₂H₃₃O₉⁺; calc. 681.2125).

Carasiphenol D (= *5,5a-Dihydro-2,7,9,12-tetrahydroxy-5,15-bis(4-hydroxyphenyl)-10H-benzo[8,9]cyclo-nona[1,2,3-c,d:4,5,6-c',d']bisbenzofuran-10-one*; **2**): Pale yellow amorphous powder. M.p. > 240°. [α]_D²⁵ = +47.9 (*c* = 0.4 MeOH). UV (MeOH): 326 (3.5). IR (KBr): 3330, 1710, 1640, 1548, 1466, 1280, 1203, 1129, 1057, 825. ¹H- and ¹³C-NMR: *Tables 1* and *2*. HR-ESI-MS: 587.1339 ([*M* + H]⁺, C₃₅H₂₃O₉⁺; calc. 587.1342).

Carasinaurone (= *2-(4-Hydroxybenzyl)-2,6-dihydroxy-4-methoxybenzofuran-3(2H)-one*; **3**): Pale yellow cubic crystals. M.p. > 240°, [α]_D²⁵ = +38.9 (*c* = 0.356, MeOH), UV (MeOH): 272 (1.7). IR (KBr): 3373, 1680, 1598, 1516, 1430, 1357, 1206, 1120, 834, 790. ¹H- and ¹³C-NMR: *Tables 1* and *2*. HR-ESI-MS: 341.0427 ([*M* + K]⁺, C₁₆H₁₄KO₆⁺; calc. 341.0427).

REFERENCES

- [1] Jiangsu New Medical College, 'The Dictionary of Traditional Medicine', Shanghai Scientific & Technical Press, Shanghai, 1975, p. 1402.
- [2] W. F. Jin, W. J. Zhu, H. F. Wang, H. F. Luo, C. Q. Hu, *Osteoporos* **2001**, *7*.
- [3] H. F. Luo, L. P. Zhang, C. Q. Hu, *J. Chin. Pharm. Sci.* **2000**, *9*, 165.
- [4] H. F. Luo, L. P. Zhang, C. Q. Hu, *Tetrahedron* **2001**, *57*, 4849.
- [5] D. Y. Ma, H. F. Luo, C. Q. Hu, *Chin. J. Chem.* **2003**, *22*, 207.
- [6] Y. Oshiteru, Y. Ueno, *Phytochemistry* **1993**, *33*, 181.
- [7] M. Khan, S. Nabi, S. Parkash, *Phytochemistry* **1986**, *25*, 1946.
- [8] H. Kobayashi, M. Namikoshi, Y. T. Yokochi, *J. Antibiot.* **1996**, *49*, 873.
- [9] B. Brady, M. G. Osullivan, *W. J. Chem. Soc.* **1989**, *9*, 1552.

Received August 4, 2004